Measurement of trace nitrate concentrations in seawater by ion chromatography with valve switching*

DU Juan (杜娟)^{1,2}, FA Yun (法芸)^{1,**}, ZHENG Yue (郑岳)^{1,3}, LI Xuebing (李学兵)⁴, DU Fanglin (杜芳林)², YANG Haiyan (杨海燕)¹

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Abstract An ion chromatographic method with a valve switching facility was developed to determine trace nitrate concentrations in seawater using two pumps, two different suppressors, and two columns. A carbohydrate membrane desalter was used to reduce the high concentrations of sodium salts in samples. In this method, trace nitrate was eluted from the concentrator column to the analytical columns, while the matrix flowed to waste. Neither chemical pre-treatment nor sample dilution was required. In the optimized separation conditions, the method showed good linearity (R>0.99) in the 0.05 and 50 mg/L concentration range, and satisfactory repeatability (RSD<5%, R=6). The limit of detection for nitrate was 0.02 mg/L. Results showed that the valve switching system was suitable and practical for the determination of trace nitrate in seawater.

Keyword: ion chromatography; valve switching; seawater; nitrate ion

1 INTRODUCTION

Determining trace nitrate concentrations in seawater is very valuable for oceanographic studies (Fukushi et al., 2009). Many methods have been used to determine nitrate concentrations, such as spectrophotometry (Finch et al., 1998; Sastry et al., 2002; Zhu, 2006), gas chromatography (GC) with electron capture detector (ECD) (Walsh, 2001), ion chromatography (IC) coupled with inductively coupled plasma mass spectrometry (ICP-MS) (Bettinelli and Spezia, 1995) and ion chromatography (IC) (Helaleh and Korenaga, 2000). These methods, however, have been used in the absence of a matrix (Walsh, 2001).

IC is the most widely used analytical method to determine ion species (Dahllöf et al., 1997; Geng et al., 2008; Haddad et al., 2008). It is an easy and straightforward way to quantify major anions, such as chloride, in seawater. The determination of trace

anions in seawater remains a challenge because of the interference of salt matrices (Wilson et al., 2011). Until now, there have been few studies on the analysis of low-level ions in seawater (Gros, 2013). Huang et al. (2000) described an IC method for determining trace analytes in seawater combined with chloride removal using a pre-concentration technique. However, in this method, the analytical column may block because of colloidal AgCl, which is produced by the co-precipitation or adsorption of analytes, and/ or Ag⁺ release, in the Ag⁺ cartridges used for chloride removal (Haddad and McTaggart, 1991; Slingsby and Pohl, 1996).

¹Public Laboratory, Qingdao Institute of Bioenergy and Bioprocess Technology, Chinese Academy of Sciences, Qingdao 266101, China

² College of Materials Science and Engineering, Qingdao University of Science & Technology, Qingdao 266042, China

³ College of Food Science and Engineering, Shandong Agricultural University, Tai'an 271018, China

⁴ Heterogeneous Catalysis Conversion Group, Qingdao Institute of Bioenergy and Bioprocess Technology, Chinese Academy of Sciences, Qingdao 266101, China

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^{**} Corresponding author: fayun@qibebt.ac.cn

In recent years, the valve switching technique has been shown to be a powerful analytical tool to deal with matrix interference (Bruno et al., 2003). This technology was previously described as an effective matrix elimination method rather than a sample pretreatment method (Villasenor, 1991), and was not only applied in liquid chromatography (Opiteck et al., 1998; Wang et al., 2009), but also in ion chromatography (Huang et al., 2000; Wang et al., 2012). IC coupled with valve switching has earned a reputation for high sensitivity and automated operation because of its ability to use different stationary phases in different columns to switch and trap target ions, and eliminate matrix interference.

In this study, we developed a valve switching IC method. Trace nitrate was eluted from the concentrator column to the analytical columns, while the bulk of the matrix flowed to waste. Compared with conventional valve switching methods, this method has the advantage of using a much larger capacity suppressor with the ability to exchange more sodium salts in seawater. This means that it can eliminate matrix interference more efficiently. This is in contrast to most of the existing methods, which cannot suppress a sufficiently large quantity of the matrix in small capacity suppressors. Under optimized conditions, the method showed good linearity (R>0.99) in the 0.05 and 50 mg/L concentration range, and satisfactory repeatability (RSD<5%, n=6).

2 METHODOLGY

2.1 Instrumentation and chromatography conditions

The experiment was performed on an ICS-5000 (Thermo Scientific, Waltham, MA, USA) equipped with dual pumps, an auto-sampler, one conductivity detector, two valves (two six-port valves) and a sample loop (50 μL). 20 mmol/L sodium hydroxide (NaOH) was manually prepared for primary separation on one Ion Pac AG11-HC guard column (50 mm×4 mm, Thermo Scientific) and one Ion Pac AS11-HC column (250 mm×4 mm, Scientific), coupled with a Carbohydrate Membrane Desalter CMD 4 mm (Thermo Scientific). The other Ion Pac AG11-HC guard column (50 mm×4 mm, Thermo Scientific) was used as a concentrator column. The 20-mmol/L potassium hydroxide (KOH) mobile phase was generated by a Reagent-Free Controller (Thermo Scientific). Nitrate was eluted on the concentrator column by an eluent generator, and was separated on the other Ion Pac AS11-HC column (250 mm×4 mm, Thermo Scientific) coupled with an ASRS 300 (4 mm) (Thermo Scientific). All the instrument control and data acquisition were performed by Chromeleon 6.8 chromatography software (Thermo Scientific). The eluent flow rate was 1.0 mL/min. The column temperature was 30°C.

2.2 Preparation of standards and samples

All the solutions were prepared using 18 $M\Omega$ water (Millipore, Mosheim, France), and filtered through a 0.22-µm nylon membrane filter. Sodium hydroxide (50% w/w, certified grade) was obtained from Acros Organics (Fair Lawn, NJ, USA), and the anion standards were supplied by the National Institute of Metrology, China. Artificial seawater was prepared using sea salts and water. Seawater samples were collected from Jiaozhou Bay (Qingdao, China) and filtered through a membrane filter (pore size 0.22 µm) prior to IC analysis.

2.3 Experimental procedure

All the procedures of the system were carried out using valves and pumps interconnected by a narrow poly (ether-ether-ketone) tubing system (Fig.1). Step 1 represents sample loading. Step 2 is the sample injection stage, where chloride is first eluted from column 1 and then eliminated. Step 3 is the extracting phase, in which nitrate is extracted on the trap column. Step 4 involves analysis and re-equilibration, in which nitrate is separated on column 2. Other compounds were eluted to waste from column 1. Once the system reaches re-equilibration, the cycle restarts from the beginning.

2.4 Method validation

The method was validated for precision, linearity and repeatability. Repeatability (the percent of the relative standard deviation) was tested using six 6.0 mg/L standard replicates. Linearity calibration curves were calculated by plotting the peak areas and concentrations of the external standards. Recovery was determined by 1.0-, 6.0-, and 10.0-mg/L spiked standards.

3 RESULT AND DISCUSSION

3.1 The switching window

We used a standards mixture (18 158 mg/L chloride and 6.0 mg/L nitrite) to determine the switching and duration times for valve 2 at which recoveries were

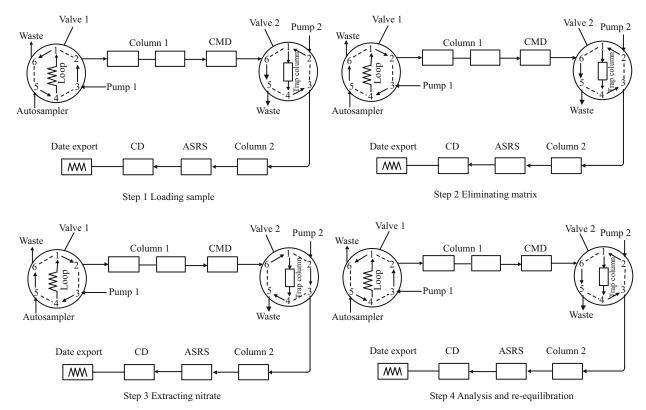


Fig.1 System coupled with valve switching

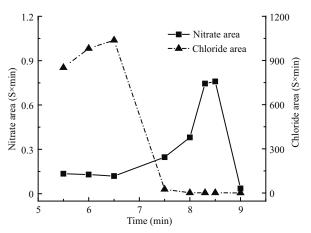
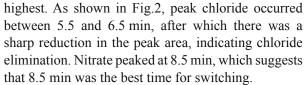


Fig.2 Switching time of valve 2

The black solid line indicates nitrate peak area (left axis); the black short dashed-dot line indicates the chloride peak area (right axis).



The duration time is shown in Fig.3, which indicates a peak value for nitrate at 2.0 min. Therefore, the optimal switching and duration times of valve 2 were 8.5 min and 2.0 min, respectively. The valve

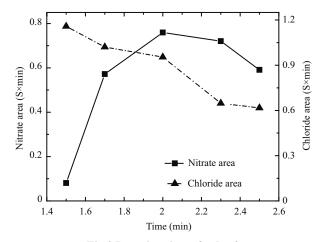


Fig.3 Duration time of valve 2

The solid black line indicates the nitrate peak area (left axis); the short black dashed-dot line indicates the chloride peak area (right axis).

switching procedure is as follows: Step 1 (-4.5–0 min), Step 2 (0–8.5 min), Step 3 (8.5–10.5 min) and Step 4 (10.5–30 min).

3.2 Method validation and sample determination

This labor-saving technology has provided us with a low cost, automated routine analysis method. This method proved easier to manipulate than using

Table 1 Reproducibility, correlation coefficient, linear range, limit of detection and average recovery

Analyte	RSD (%) (<i>n</i> =6)	Correlation coefficient (<i>n</i> =7)	Regression equation	Linear range (mg/L)	LOD (mg/L)	Average recovery (%)
Nitrate	4.830	0.999 2	Y=0.1403X-0.1057	0.05-50	0.02	99.2

monolithic ODS columns coated with 5 mmol/L cetyltrimethylammonium chloride (CTAC) aqueous solution (Ito et al., 2005). The linearity of the response was tested by 50-µL injections of standards with the following concentrations: 0.05, 0.1, 0.5, 2, 6, 10, and 50 mg/L. Results are given in Table 1. The LOD of 0.02 ppm for nitrate was lower than that achieved by an alternative column-switching method (Bruno et al., 2003). The correlation coefficient of the calibration reached 0.999 2. Repeatability for the six 6.00 mg/L standard replicates was 4.83%. The mean recovery for the three replicates of 1.0, 6.0, and 10.0 mg/L spiked standards was 99.2%. The results are more precise than adding a fixed volume of a nitrate standard solution to all the samples (Raessler and Hilke, 2006), because there are no dilution effects.

Trace nitrate in seawater was determined in optimal conditions. The IC chromatogram is displayed in Fig.4, which shows a stable baseline separation without matrix interference. From three replicates, the seawater nitrate concentration was $0.4697\pm0.001~\text{mg/L}$.

4 CONCLUSION

This study demonstrates that a single valve switching system after injection can be used successfully for the analysis of low-level nitrate. Nitrate and a small amount of chloride were obtained after the initial separation in an analytical column. Low-level nitrate was eluted from the concentrator column to the second analytical column, and the majority of the matrix was eliminated by a large capacity desalter. The proposed method has the potential to become a convenient and practical tool for routine analysis of seawater.

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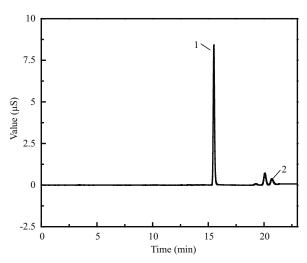


Fig.4 Chromatogram of seawater derived from the valve switching technique

1=chloride: 2=nitrate.

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