

Evaluation of the separation performance of polyvinylpyrrolidone as a virtual stationary phase for chromatographic NMR

Shaohua Huang,^{a*} Rui Wu,^b Zhengwu Bai,^{b**} Ying Yang,^a Suying Li^a and Xiaowei Dou^a

Polyvinylpyrrolidone (PVP) was used as a virtual stationary phase to separate *p*-xylene, benzyl alcohol, and *p*-methylphenol by the chromatographic NMR technique. The effects of concentration and weight-average molecular weight (Mw) of PVP, solvent viscosity, solvent polarity, and sample temperature on the resolution of these components were investigated. It was found that both higher PVP concentration and higher PVP Mw caused the increase of diffusion resolution for the three components. Moreover, the diffusion resolution did not change at viscosity-higher solvents. Moreover, the three components showed different resolution at different solvents. As temperature increased, the diffusion resolution between *p*-xylene and benzyl alcohol gradually increased, and the one between *p*-xylene and *p*-methylphenol slightly increased from 278 to 298 K and then decreased above 298 K. It was also found that the polarity of the analytes played an important role for the separation by affecting the diffusion coefficient. Copyright © 2014 John Wiley & Sons, Ltd.

Keywords: NMR; ¹H; chromatographic NMR; polyvinylpyrrolidone; virtual stationary phase; diffusion resolution

Introduction

Diffusion-ordered spectroscopy (DOSY) in NMR is a new effective tool for the analyses of complicated mixture.^[1–4] In DOSY experiment, the diffusion coefficients (*D*) related to separated components are plotted on the vertical axis (F1), while the chemical shifts in the spectrum yielded by each component are plotted on the horizontal axis (F2).^[5] Herein, *D* is described by the Stokes–Einstein equation:

$$D = \frac{kT}{6\pi\eta r_s}$$

where, *k* is the Boltzmann constant, *T* refers to the temperature, η represents the viscosity of the liquid, and r_s is the (hydrodynamic) radius of the molecule. From the Stokes–Einstein equation, it is obvious that *T*, η , and r_s are the three important factors.

The application of DOSY has been dubbed chromatographic NMR (CNMR),^[4,6–8] which can be evaluated by the diffusion resolution with a formula:

$$\Delta D = D_{A1} - D_{A2}$$

where, D_{A1} is the *D* of analyte 1 (A1) and D_{A2} is the *D* of analyte 2 (A2), respectively. *D* depends on many physical parameters such as mass, size, and shape of a molecule, sample temperature, and viscosity of analyses system. Therefore, a complicated mixture can be fully separated as long as the differences in some physical parameters of each component are prominent enough. However, it is not easy to obtain a desirable resolution only through the pulsed field gradient NMR technique, especially for these components which bear similar molecular properties. A practical approach to solve this

problem is adding some polymer,^[9–11] surfactant,^[12–14] microemulsion,^[15–19] or silica gel^[3,20–26] in NMR tube or NMR rotor as a matrix or a virtual stationary phase (VSP) to enlarge the difference of diffusion rates of the components. DOSY techniques can discriminate complex components very well by using these additives. Up to now, there is no report concerning the influences of experimental conditions on the resolution. In order to further understand the mechanism of CNMR and to enlarge the application scope of this technique, it is of significance to observe the influence of these conditions such as VSP type, solvent viscosity, solvent polarity, and sample temperature on the separation. In 2009, Kavakka J. S. *et al.*^[9] reported that PVP (refer to Fig. 1) can fully separate a mixture of *p*-xylene, benzyl alcohol, and *p*-methylphenol by ¹H CNMR technique, but the effect of ‘chromatographic conditions’ on diffusion resolution was not studied. In this work, we employed PVP as a VSP and evaluated its separation performance in ¹H CNMR under different conditions including concentration and Mw of PVP, solvent viscosity, solvent polarity, and sample temperature.

* Correspondence to: Shaohua Huang, Key Laboratory of Biobased Materials, Qingdao Institute of Bioenergy and Bioprocess Technology, Chinese Academy of Sciences, Qingdao 266101, China. E-mail: huangsh@qibebt.ac.cn

** Correspondence to: Zhengwu Bai, School of Chemistry and Environmental Engineering, Wuhan Institute of Technology, Wuhan 430073, China. E-mail: zhengwu_bai@yahoo.com

a Key Laboratory of Biobased Materials, Qingdao Institute of Bioenergy and Bioprocess Technology, Chinese Academy of Sciences, Qingdao 266101, China

b School of Chemistry and Environmental Engineering, Wuhan Institute of Technology, Wuhan 430073, China

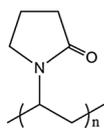


Figure 1. Structure of PVP.

Experimental

Chemicals and instruments

PVPs with different Mws ($M_w = 10\,000$, $24\,000$, $58\,000$ g/mol) were purchased from Chengdu Ai Keda Chemical Technology Co., Ltd. (China) and used without pre-treatment. *p*-Xylene, benzyl alcohol, *p*-methylphenol, and CCl_4 were all obtained from Sinopharm Chemical Reagent Co., Ltd. (China) and used as received. CDCl_3 and CD_3OD were purchased from Cambridge Isotope Laboratories, Inc. (USA) with 0.003% (v/v) TMS as internal standard. CNMR measurements were conducted on a high-resolution liquid NMR 600 MHz spectrometer of Bruker Avance III (Sweden) with a 5 mm TCI CryoProbe equipped with Z-gradients up to 53 g/cm operating at 600.13 MHz.

NMR experiments

In all CNMR experiments, the DOSY data sets were measured using four steady-state scans, eight transients, and 1.5 s relaxation delay. Spectral width in ^1H -dimension was 7211.539 Hz, and the number of acquired complex data points was 4096. Diffusion was measured using a LEDBPGPCPMG2S pulse sequence, ramping the strongest gradient from 2 to 95% of maximum strength in 30 steps. Spoil gradient duration was 0.6 ms, and eddy current recovery delay was 5 ms. Diffusion gradient duration and diffusion time were given according to the practical needs. The duration of Carr–Purcell–Meiboom–Gill (CPMG)-filter of 1 ms was used, and the total length of CPMG-train for polymer signal suppression was 60 ms. Sample temperature was set at 298 K except when indicated. The data was apodized by sine function and zero-filled up to 16 384 complex points prior to Fourier transformation. The 2D DOSY data set was processed by using the command DOSY2D in Bruker TopSpin 3.1 spectrometer operating software. Notably, bipolar longitudinal eddy current delay with gradients (LEDBPGP^[27]) pulse sequence was modified in order to make this sequence include a T2-filter element (CPMG spin echo train, CPMG^[28,29]) prior to the acquisition period. This modified pulse sequence was named as LEDBPGPCPMG2S shown in Fig. 2.

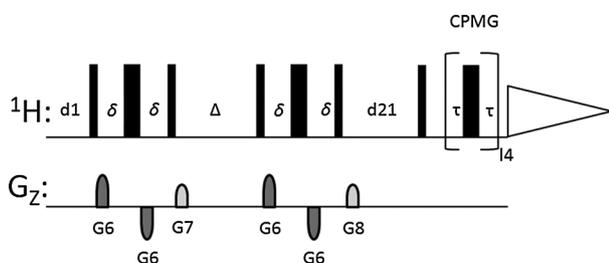


Figure 2. Pulse sequence for LEDBPGPCPMG2S.

Results and Discussion

In CNMR experiments, the additive structure should be preferably simple, avoiding the possible overlap with analyte signals. In addition, VSP must not cause too high viscosity for the solvent, otherwise resulting in line broadening of analyte signals. Based on this consideration and the reported observations,^[9] PVP was selected as the VSP and a mixture of *p*-xylene, benzyl alcohol, and *p*-methylphenol as the analyte in this work. CDCl_3 was used as the NMR solvent to form a 'normal-phase' system because the molecular polarity of PVP is higher than that of CDCl_3 . Besides, pulse program with T_2 filtering in CNMR measurements was loaded in order to suppress the VSP signals. Under these conditions, the three components in the mixture were not separated in DOSY spectra in the absence of PVP (Fig. 3(a)), and on the contrary, *p*-xylene, benzyl alcohol, and *p*-methylphenol were satisfactorily resolved in the presence of PVP (Fig. 3(b)). The result of this CNMR measurement demonstrates that PVP is a preferable VSP for these three components.

Influence of PVP concentration on diffusion coefficient and resolution

Figure 4 shows the influence of PVP concentration (C) on D and ΔD values. As seen in Fig. 4, D values of three components became small as PVP concentration increased. Moreover, the absolute value of the slope (designed as S) of the reduction in D values of these three components is in a sequence of $S_{p\text{-xylene}} > S_{\text{benzyl alcohol}} > S_{p\text{-methylphenol}}$. This sequence correlates the component polarity, which is in a sequence of *p*-xylene < benzyl alcohol < *p*-methylphenol. As shown in Fig. 4, ΔD_1 ($D_{p\text{-xylene}} - D_{\text{benzyl alcohol}}$) and ΔD_2 ($D_{p\text{-xylene}} - D_{p\text{-methylphenol}}$) were enlarged as PVP concentration increased. The slopes of the increase in ΔD_1 and ΔD_2 values were higher when C was lower than 25 mg/ml, revealing that the components were resolved better at the higher concentrations of PVP. However, PVP limitedly improved the resolutions when C was higher than 25 mg/ml.

In fact, when PVP was fed at an increasing amount, more structural units of *N*-vinylpyrrolidone in the solvent interacted with the components causing the increases of ΔD . On the other hand, more PVP brought about a higher viscosity of the solvent, and as a result, the diffusion of the components was restricted. This trend was evidenced by the decrease of D . The increase of ΔD was accordingly limited when PVP was fed at a concentration above 25 mg/ml.

Influence of PVP Mw on diffusion coefficient and resolution

Another important factor impacting D and ΔD is the Mw of VSP. In the present work, three PVPs of different Mws were exploited as VSPs. The dependence of D and ΔD on the Mws is presented in Fig. 5. As shown in Fig. 5, D values of three components reduced as the Mw of PVPs increased and decreased very slowly when the Mw was more than 24 000 g/mol. Two ΔD values were enlarged as the increase of Mw and were almost invariable when Mw was between 24 000 and 58 000 g/mol. The weight of PVPs of different Mw was fed by 50 mg in 0.6 ml CDCl_3 . In each experiment, the number of structural units was same for each VSP. Therefore, the changes in D and ΔD values were created by the different Mw of the polymers. The chain of PVPs of higher Mw twins hinders the motion of the components and further reduces ΔD . This observation demonstrates that a certain amount of Mw

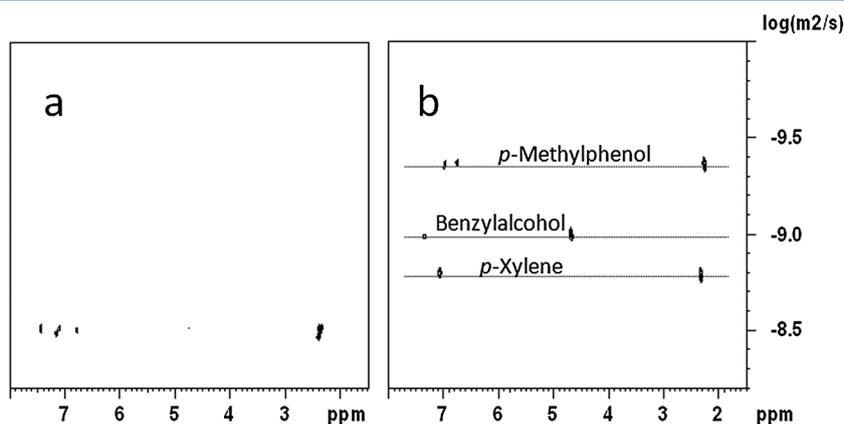


Figure 3. ^1H NMR spectra (600 MHz) of mixture including *p*-xylene (20 mg), benzyl alcohol (20 mg), and *p*-methylphenol (20 mg) before (a) and after (b) adding 50 mg PVP (Mw = 10 000 g/mol) in 0.6 ml CDCl_3 when T was set at 298 K.

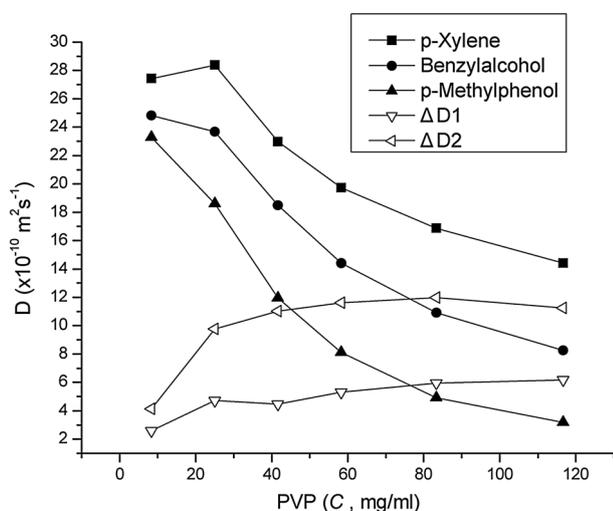


Figure 4. Influence of PVP concentration (C) on diffusion coefficient and resolution. PVP Mw: 10 000 g/mol; each component: 20 mg; solvent: 0.6 ml CDCl_3 ; T : 298 K.

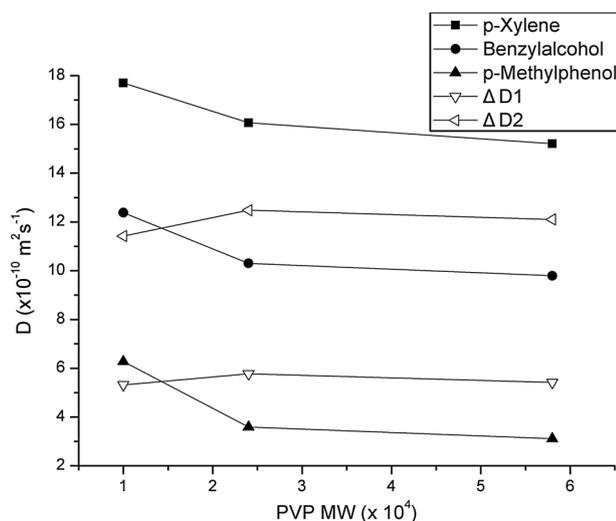


Figure 5. Influence of PVP Mw on diffusion coefficient and resolution. PVP: 50 mg; each component: 20 mg; solvent: 0.6 ml CDCl_3 ; T : 298 K.

is necessary for a VSP to separate components of a sample. However, higher Mw may not be favorable for better resolution.

Influence of solvent viscosity and polarity on diffusion coefficient and resolution

Solvent is an important element impacting experiment results in chromatography, especially in liquid chromatography and thin layer chromatography.^[30–33] Solvent properties such as viscosity and polarity etc., influence the existing state of a sample molecule in solvent and further affect the retention properties such as retention factor, selectivity, and resolution. Similarly, the solvents in CNMR may also affect separation performance of a VSP.

When the measurement was conducted, CDCl_3 was employed as the solvent and CCl_4 as a viscosity modifier. The viscosity of CDCl_3 is 0.54 mPa s and is 0.91 mPa s for CCl_4 at 25 °C. The effect of solvent viscosity on D and ΔD values is presented in Fig. 6. As shown in Fig. 6, D values of three components reduced as the increase of CCl_4 in solvent. Obviously, the solvent of higher viscosity impedes the diffusion of components, resulting in the decrease in D values of the components. Additionally, D value may be related to the polarity of the components. D value is in

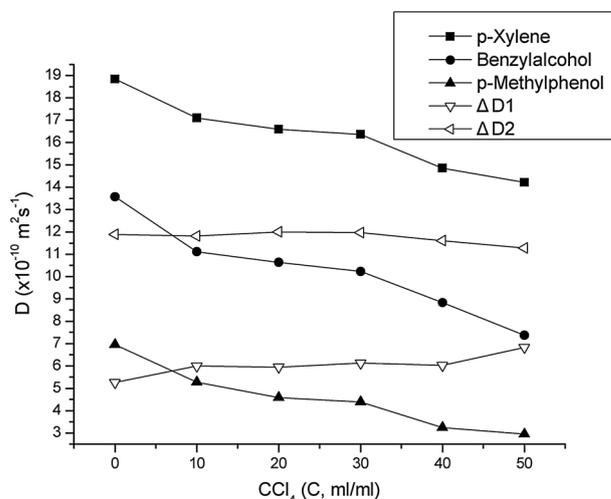


Figure 6. Influence of solvent viscosity on diffusion coefficient and resolution. PVP Mw: 10 000 g/mol; PVP: 50 mg; each component: 5 mg; solvent volume: 0.6 ml; T : 298 K.

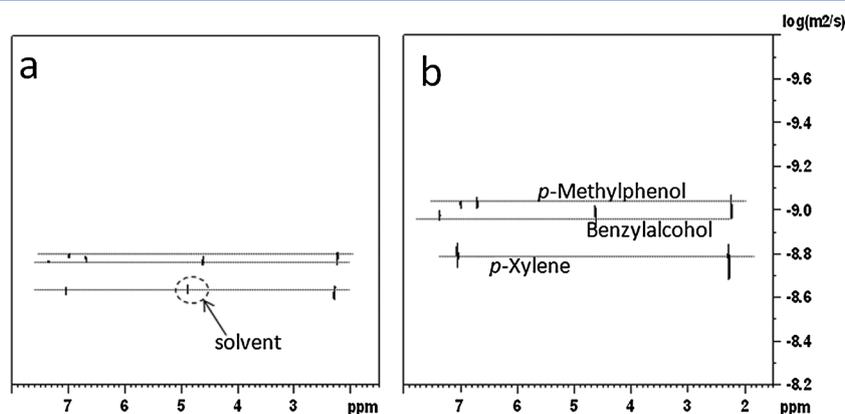


Figure 7. ^1H CNMR spectra (600 MHz) of mixture including *p*-xylene (20 mg), benzyl alcohol (20 mg), and *p*-methylphenol (20 mg) before (a) and after (b) adding 50 mg PVP (Mw = 10 000 g/mol) in 0.6 ml CD_3OD when T was set at 298 K.

a sequence of $D_{p\text{-xylene}} > D_{\text{benzyl alcohol}} > D_{p\text{-methylphenol}}$, while the polarity is in a sequence of $p\text{-xylene} < \text{benzyl alcohol} < p\text{-methylphenol}$. The two reversed sequences imply that the component of lower polarity shows higher diffusion. In the measurements, the solvent was CDCl_3 or a mixture of CDCl_3 and CCl_4 . Chloro group is an electron-rich substituent. The hydroxyls in *p*-methylphenol and benzyl alcohol interacted with the chloro groups of the solvents, decreasing the diffusion of the two components. Because the acidity of *p*-methylphenol is stronger than that of benzyl alcohol, *p*-methylphenol interacts stronger with chloro group than benzyl alcohol. Therefore, the D value of *p*-methylphenol should be lower than that of benzyl alcohol, and *p*-xylene should have a highest D value. This inference is evidenced by Figs. 4–6 and 8.

On the other hand, although D values varied, ΔD values almost remained unchanged as the increase of solvent viscosity, indicating that solvent viscosity affects D values but does not influence the resolution in these CNMR detections. The reason should be that the interaction between PVP and the components is stronger than that between solvent and the components, and the resolution (ΔD values) is dominated by the stronger interaction.

The effect of the solvent polarity on resolution was observed through the experiments in different solvents. Figure 7 shows the ^1H DOSY spectra of *p*-xylene, benzyl alcohol, and *p*-methylphenol in CD_3OD without and with PVP, respectively. As seen in Fig. 7(a), benzyl alcohol and *p*-methylphenol were not fully separated from each other, while *p*-xylene was resolved from the above two components in the absence of PVP; on the contrary, the three components were fully separated in the presence of PVP (Fig. 7(b)). The observation in CD_3OD is different from that in CDCl_3 , where the three components had not been separated from each other in the absence of PVP. Therefore, solvent polarity not only influences the separation result of pure DOSY experiment but also changes the separation performance of CNMR.

Influence of sample temperature on diffusion coefficient and resolution

Temperature usually affects chromatographic separation,^[34–36] and similarly may impact CNMR measurements because molecular translational motion closely depends on sample temperature. In this work, the effect of experimental temperature on D and ΔD values of three components was investigated. Figure 8 shows the

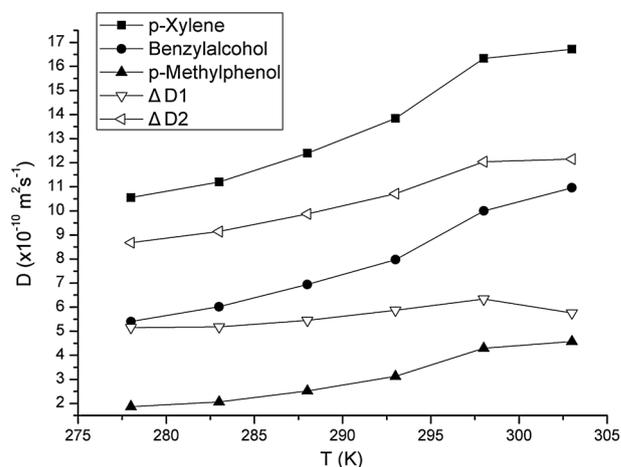


Figure 8. Influence of sample temperature (T) on diffusion coefficient and resolution. PVP Mw: 10 000 g/mol; PVP: 50 mg; each component: 20 mg; solvent: 0.6 ml CDCl_3 .

variation in D values of three components along of the increase of temperature. The slope of the increase in D value of the three components is in a sequence of $S_{p\text{-xylene}} > S_{\text{benzyl alcohol}} > S_{p\text{-methylphenol}}$, which is reversed to the polarity sequence of $p\text{-xylene} < \text{benzyl alcohol} < p\text{-methylphenol}$. This trend reveals that temperature more significantly affects the D value of lower polar molecule and comparatively less affects that of higher polar molecule. With respect to the variation of ΔD , ΔD_1 increased slightly from 278 to 298 K, then decreased above 298 K. While ΔD_2 almost increased as the temperature rose. The variation trend of ΔD_1 and ΔD_2 is different when the sample temperature is above 298 K.

Understanding the mechanism of resolution

Stokes–Einstein equation shows that T , η , and r_s are the three significant factors to affect D value. The experimental results in the present work, however, reveal that molecular polarity significantly influences the ΔD values of three components in CNMR measurement. The different molecular polarity of the components causes the different strength of the interaction between the components and the VSP, leading to the change in D and ΔD values. Therefore, the polarity of the components should principally impact the separation of the components.

Conclusions

The effects of PVP concentration, PVP Mw, solvent viscosity, solvent polarity, and sample temperature on the ΔD values of *p*-xylene, benzyl alcohol, and *p*-methylphenol were investigated by the ^1H CNMR technique, and these 'chromatographic conditions' affected the ΔD values. As the increase of PVP concentration and PVP Mw, ΔD values increased. When the solvent consisted of CDCl_3 and CCl_4 , the increased solvent viscosity did not change the ΔD values, while the separation result in CDCl_3 was different from that in CD_3OD . In addition, ΔD_2 increased along of the increase of temperature. While ΔD_1 firstly slightly increased, and then decreased. ΔD_1 was different from ΔD_2 in the variation trend when the sample temperature is above 298 K. Moreover, the molecular polarity of the components principally impacted the diffusion behavior and the separation for this 'normal-phase' CNMR.

Acknowledgement

The financial support from National Natural Science Foundation of China (21105108) is gratefully acknowledged.

References

- [1] K. F. Morris, P. Stilbs, C. S. Johnson. *Anal. Chem.* **1994**, *66*, 211.
- [2] C. Carrara, C. Lopez, S. Caldarelli. *J. Chromatogr. A* **2012**, *1257*, 204.
- [3] C. Carrara, S. Caldarelli. *J. Phys. Chem. C* **2012**, *116*, 20030.
- [4] R. Novoa-Carballal, E. Fernandez-Megia, C. Jimenez, R. Riguera. *Nat. Prod. Rep.* **2011**, *28*, 78.
- [5] C. S. Johnson. *Prog. Nucl. Magn. Reson. Spectrosc.* **1999**, *34*, 203.
- [6] S. Caldarelli. *Magn. Reson. Chem.* **2007**, *45*, S48.
- [7] K. E. Price, L. H. Lucas, C. K. Larive. *Anal. Bioanal. Chem.* **2004**, *378*, 1405.
- [8] J. S. Gounarides, A. D. Chen, M. J. Shapiro. *J. Chromatogr. B* **1999**, *725*, 79.
- [9] J. S. Kavakka, I. Kilpelainen, S. Heikkinen. *Org. Lett.* **2009**, *11*, 1349.
- [10] J. S. Kavakka, V. Parviainen, K. Wahala, I. Kilpelainen, S. Heikkinen. *Magn. Reson. Chem.* **2010**, *48*, 777.
- [11] R. E. Joyce, I. J. Day. *J. Magn. Reson.* **2012**, *220*, 1.
- [12] J. Cassani, M. Nilsson, G. A. Morris. *J. Nat. Prod.* **2012**, *75*, 131.
- [13] C. F. Tormena, R. Evans, S. Haiber, M. Nilsson, G. A. Morris. *Magn. Reson. Chem.* **2010**, *48*, 550.
- [14] M. E. Zielinski, K. F. Morris. *Magn. Reson. Chem.* **2009**, *47*, 53.
- [15] C. Pemberton, R. E. Hoffman, A. Aserin, N. Garti. *Langmuir* **2011**, *27*, 4497.
- [16] R. Evans, S. Haiber, M. Nilsson, G. A. Morris. *Anal. Chem.* **2009**, *81*, 4548.
- [17] K. F. Morris, B. A. Becker, B. C. Valle, I. M. Warner, C. K. Larive. *J. Phys. Chem. B* **2006**, *110*, 17359.
- [18] B. A. Begotka, J. L. Hunsader, C. Oparaache, J. K. Vincent, K. F. Morris. *Magn. Reson. Chem.* **2006**, *44*, 586.
- [19] S. R. Chaudhari, N. Suryaprakash. *J. Mol. Struct.* **2012**, *1017*, 106.
- [20] C. Pemberton, R. Hoffman, A. Aserin, N. Garti. *J. Magn. Reson.* **2011**, *208*, 262.
- [21] J. Xu, T. W. Tan, L. Kenne, C. Sandstrom. *New J. Chem.* **2009**, *33*, 1057.
- [22] R. E. Hoffman, H. Arzuan, C. Pemberton, A. Aserin, N. Garti. *J. Magn. Reson.* **2008**, *194*, 295.
- [23] C. Carrara, S. Viel, C. Delaurent, F. Ziarelli, G. Excoffier, S. Caldarelli. *J. Magn. Reson.* **2008**, *194*, 303.
- [24] G. Pages, C. Delaurent, S. Caldarelli. *Angew. Chem. Int. Ed.* **2006**, *45*, 5950.
- [25] S. Viel, F. Ziarelli, S. Caldarelli. *Proc. Natl. Acad. Sci. U. S. A.* **2003**, *100*, 9696.
- [26] G. N. M. Reddy, R. Ballesteros-Garrido, J. Lacour, S. Caldarelli. *Angew. Chem. Int. Ed.* **2013**, *52*, 1.
- [27] D. H. Wu, A. D. Chen, C. S. Johnson. *J. Magn. Reson., Ser. A* **1995**, *115*, 260.
- [28] H. Y. Carr, E. M. Purcell. *Phys. Rev.* **1954**, *94*, 630.
- [29] S. Meiboom, D. Gill. *Rev. Sci. Instrum.* **1958**, *29*, 688.
- [30] F. Zhan, G. Yu, B. Yao, X. Guo, T. Liang, M. Yu, Q. Zheng, W. Weng. *J. Chromatogr. A* **2010**, *1217*, 4278.
- [31] K. Le Mapihan, J. Vial, A. Jardy. *Chromatographia* **2003**, *57*, S163.
- [32] U. D. Neue, C. H. Phoebe, K. Tran, Y. Cheng, Z. Lu. *J. Chromatogr. A* **2001**, *925*, 49.
- [33] J. Barbosa, I. Toro, R. Berges, V. Sanz-Nebot. *J. Chromatogr. A* **2001**, *915*, 85.
- [34] G. Paglia, O. D'Apolito, F. Tricarico, D. Garofalo, G. Corso. *J. Sep. Sci.* **2008**, *31*, 2424.
- [35] Y. Yang. *Anal. Chim. Acta* **2006**, *558*, 7.
- [36] D. Zhang, F. Li, D. H. Kim, H. J. Choi, M. H. Hyun. *J. Chromatogr. A* **2005**, *1083*, 89.