



Efficiency of CO₂ fixation by microalgae in a closed raceway pond



Shuwen Li^{a,b,c}, Shengjun Luo^{a,*}, Rongbo Guo^{a,*}

^a Key Laboratory of Biofuels, Qingdao Institute of Bioenergy and Bioprocess Technology, Chinese Academy of Sciences, Qingdao 266101, PR China

^b College of Life Sciences, Qingdao Agricultural University, Qingdao 266109, PR China

^c University of Chinese Academy of Sciences, Beijing 100049, PR China

HIGHLIGHTS

- A closed raceway pond and its equation of CO₂ mass transfer.
- CO₂ dissolution efficiency in the closed raceway pond without algae.
- CO₂ fixation efficiency by algae with continuous aeration in the pond.
- A mode of intermittent aeration with CO₂ fixation efficiency of 95% in the pond.

ARTICLE INFO

Article history:

Received 3 December 2012

Received in revised form 4 March 2013

Accepted 6 March 2013

Available online 13 March 2013

Keywords:

Microalgae

Fixation efficiency

Mass transfer coefficient

Model

Intermittent gas sparging

ABSTRACT

Microalgae contain about 50% of carbon, which means that a total of 1.83 ton of CO₂ is needed to produce 1 ton of microalgae. The cost of CO₂ supply for microalgal large scale cultivation should be considered and the low CO₂ fixation efficiency by microalgae will lead to much more expenditure of CO₂. In this study, a closed raceway pond was constructed by covering a normal open raceway pond with a specially designed transparent cover, which directly touched the surface of microalgal culture media. This cover prevented supplied CO₂ escaping into atmosphere and thus increased the retention time of CO₂. The CO₂ gas–liquid mass transfer and CO₂ fixation efficiency by microalgae in the closed raceway pond were investigated, and the model of CO₂ fixation by microalgae was developed. Through the model, the CO₂ fixation efficiency increased to 95% under intermittent gas sparging.

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

Microalgae have received extensive research for five decades because of their possible commercial application in biofuels, cosmetics, pharmaceuticals, nutrition and food additives, aquaculture, pollution prevention, wastewater treatment and so on (Mata et al., 2010; Milledge, 2010; Spolaore et al., 2006). However, the high cost of microalgal cultivation is one of the greatest obstacles for commercial application. The availability and cost of CO₂ are major factors in the economics of algal mass culture. It was estimated that the cost of carbon source ranged from about 8% to 27% of the production costs of the algae (Laws and Berning, 1991). The cost of carbon source will be multiplied with the lower CO₂ fixation efficiency. There are many factors influencing CO₂ fixation efficiency in microalgal cultivation, including microalgal strains, structures of photobioreactors, light quantity and quality, operating models and nutrition conditions. The structures of photobiore-

actors have an important effect on CO₂ fixation efficiency because they affect CO₂ dissolution efficiency and CO₂ utilization efficiency by microalgae. Open-culture systems normally cannot use supplied CO₂ effectively, which easily escaped from the culture media due to the shallow depths and poor CO₂ mass transfer efficiency, and CO₂ fixation efficiency ranged from 10% to 30% (Becker, 1994; Weissman and Goebel, 1985). This low efficiency results in microalgal cultivation cost intensive (Campbell et al., 2011). In order to improve CO₂ fixation efficiency by microalgae in open raceway pond, Ketheesan and Nirmalakhandan (2012) designed an airlift-driven raceway reactor for microalgal cultivation with the maximum CO₂ utilization efficiency of 33%, and Putt et al. (2011) designed and investigated CO₂ transfer efficiency of a carbonation column combining with open ponds without analyzing the energy consumption and the microalgal productivity in the photobioreactor. Compared with open culture systems, closed photobioreactors could reduce CO₂ losses because they can prolong CO₂ retention time and improve mass transfer efficiency. Many researchers have studied microalgal CO₂ fixation with closed photobioreactors (Chiu et al., 2008; de Morais and Costa, 2007; Cheng et al., 2006; Keffer and Kleinheinz, 2002). But the CO₂ fixation efficiency was not high,

* Corresponding authors. Tel./fax: +86 532 80662750 (S. Luo), tel.: +86 532 80662708; fax: +86 532 80662709 (R. Guo).

E-mail addresses: luosj@qibebt.ac.cn (S. Luo), guorb@qibebt.ac.cn (R. Guo).

especially for the high concentration of CO₂. Chiu et al. (2008) reported that the efficiency of CO₂ removal by *Chlorella* sp. was 16% at CO₂ concentration of 15%, and in treatment of 2% CO₂ gas sparging, CO₂ fixation efficiency was 58% in a semi-continuous photobioreactor. de Moraes and Costa (2007) reported that the CO₂ fixation efficiency was lower with high CO₂ concentration (12%) than that with low concentration (6%). In the present study, a closed raceway pond was proposed and its structure can increase the CO₂ retention time. Cheng et al. (2006) had demonstrated that increasing retention time of CO₂ in photobioreactor will significantly enhance the CO₂ fixation efficiency. The objective of this work was to investigate the CO₂ fixation efficiency by microalgae at high CO₂ concentration in the closed raceway pond and develop dynamic model of CO₂ fixation, which is used to provide method for the improvement of CO₂ fixation efficiency.

2. Methods

2.1. Closed raceway pond (CRWP)

The schematic diagram of the closed raceway pond is shown in Fig. 1. The CRWP was a four-channel raceway-type pond, being 1.2 m long, 0.25 m wide and 0.09 m deep, with each channel 0.06 m width. The CRWP had two parts and was made of polymethyl methacrylate. The bottom part of the CRWP was a rectangular box without top surface, and the cover part was H-shaped with the water clapboards attached to the lower surface of the cover. A paddle wheel was used for mixing and stirring of the culture medium. The enriched CO₂ gas was aerated into the CRWP by a porous tube from CO₂ inlet. As Fig. 1 shows, the H-shaped cover directly touched the culture liquid surface and kept gas bubbles going along with culture flow. The dashed line in Fig. 1 represents the surface of culture liquid, which touched the cover. The culture liquid was 0.04 m deep. The outlet of exhausted gas in CRWP was in the paddle wheel mixing zone. The arrows in channels represent the direction of culture flow.

2.2. Microalgal cultivation

Strain *Chlorella vulgaris* was obtained from College of Marine Life Sciences, Ocean University of China. Algal cells were cultivated in the following medium (per liter) (Li et al., 2012), containing 1000 mg KNO₃, 237 mg KH₂PO₄, 88 mg CaCl₂·2H₂O, 40 mg EDTA, 30 mg FeSO₄·7H₂O, 204 mg MgSO₄·7H₂O, and 1 mL of trace metal solution. The trace metal solution (per liter) included 0.83 g H₃BO₃, 0.17 g (NH₄)₆Mo₇O₂₄·4H₂O, 0.51 g CoCl₂·6H₂O, 3.3 g MnCl₂·4H₂O, 0.95 g CuSO₄·5H₂O, 2.7 g ZnSO₄·7H₂O. The flow velocity of culture medium in CRWP was 0.03 m s⁻¹ with 12 L working volume. Culture was aerated continuously (or intermittently) with 15% CO₂ at different gas flow rates, and the gas flow rate was mea-

sured using a quality flow meter (LZB-3, Yinhuan, China). The microalgal cells were incubated at 24 ± 1 °C under the light intensity of 100 μmol m⁻² s⁻¹ with continuous light, and the light intensity was determined using a light meter (Hansatech Instruments Quantitherm light meter thermometer, Norfolk, England). The pH value of culture was measured using a pH meter (PB-10, Sartorius, Germany) and the concentration of dissolved oxygen (DO) in culture was determined using a DO meter (M700, Mettler Toledo, Switzerland). The dissolved CO₂ concentration in media was determined by a CO₂ meter (FC-200, Shanghai su park information technology Co. Ltd., China).

Culture suspension with a volume of 30 mL was filtered through pre-dried and pre-weighted 2 μm membrane filters, after filtration the filter was rinsed with distilled water, and the filter with microalgal cells was dried at 105 °C in an oven overnight. The filter was removed from the oven, kept in a desiccator and reweighed. This weight minus the weight of the empty filter gave the biomass density. Samples were generally run in triplicate for each data point. The biomass productivity (P , g L⁻¹ d⁻¹) of microalgae was calculated from the variation in biomass density (X , g L⁻¹) according to the equation $P = \frac{X_t - X_0}{t - t_0}$, where X_0 was the initial biomass density at time t_0 and X_t was the biomass density at any time t subsequent to t_0 .

2.3. Calculation of CO₂ volumetric mass transfer coefficient

The CRWP containing 12 L media without microalgal cultivation was aerated with different concentrations of CO₂ (1.2%, 2%, 15%, 100% (v/v) with N₂ balance) for 120 min at 24 ± 0.5 °C. The liquid velocity (v_{wat}) was 0.03 m s⁻¹ and the dissolved CO₂ concentration in culture media was recorded every five minutes. According to two-film theory (Carvalho et al., 2006) and Henry's law, the volumetric mass transfer coefficient of CO₂ in media was calculated from the following equation:

$$\frac{dC}{dt} = K_L a (C^* - C) \quad (1)$$

$\frac{dC}{dt}$, volumetric transport rate of CO₂ in liquid (mol min⁻¹ L⁻¹); $K_L a$, volumetric mass transfer coefficient of CO₂ (min⁻¹); C^* , concentration of CO₂ in culture in equilibrium with CO₂ content in CO₂ bubbles (mol L⁻¹), which can be determined via Henry's Law; C , CO₂ concentration in culture media (mol L⁻¹).

Integration for $C = C_0$ at $t = 0$, led to the following equation:

$$\ln \frac{C^* - C}{C^* - C_0} = -K_L a t$$

A plot of left hand side of this equation against time was used to obtain $-K_L a$ as the slope.

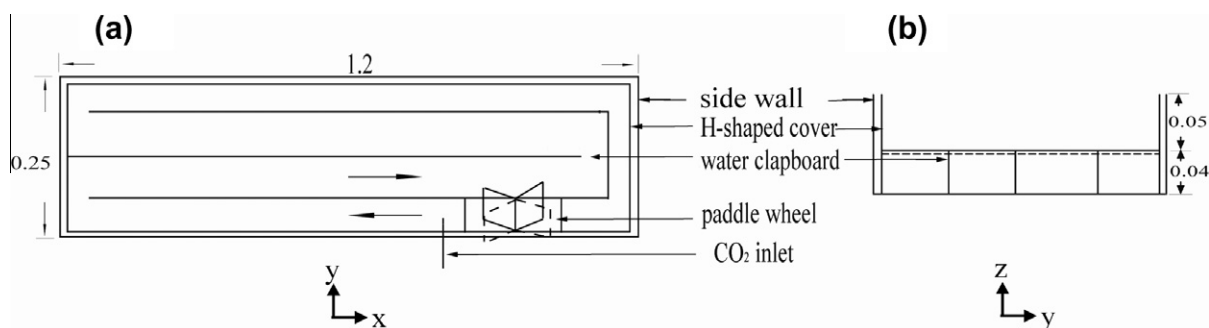


Fig. 1. The schematic diagram of the closed raceway pond (a): top view of the closed raceway pond; (b): side view of the closed raceway pond.

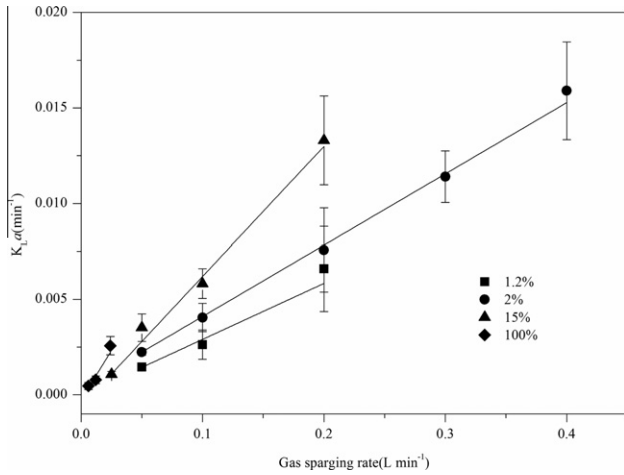


Fig. 2. The relation between gas sparging rate and K_La .

3. Results and discussion

3.1. CO₂ volumetric mass transfer and dissolution efficiency in CRWP

The CO₂ volumetric mass transfer coefficients of different concentrations of CO₂ in culture media in CRWP aerated with different rates are shown in Fig. 2. With the same gas sparging rate, K_La increased with the increase of CO₂ concentrations. Under the same CO₂ concentration, K_La was proportional to gas sparging rate. A linear regression equation between K_La and gas sparging rate (Q) can be obtained:

$$K_La = AQ \quad (2)$$

where Q is the volumetric flow rate of gas (L min⁻¹) and A is a coefficient.

The A value at CO₂ concentrations of 1.2, 2, 15, and 100% was 0.031 ($R^2 = 0.9894$), 0.039 ($R^2 = 0.9991$), 0.065 ($R^2 = 0.9944$), 0.097 ($R^2 = 0.9568$), respectively. Integration of Eqs. (1) and (2) led to:

$$\frac{dC}{dt} = A(C^* - C)Q \quad (3)$$

The transport rate of CO₂ in CRWP aerated with different gas flow rates and CO₂ concentrations can be calculated from Eq. (3).

According to Eq. (3), when the change of $(C^* - C)$ can be neglected in short time, the number of mole of dissolved CO₂ (n_{CO_2-dis} , mol) in culture media can be expressed as follows:

$$n_{CO_2-dis} = A(C^* - C)QtV$$

where t is the gas sparging time (min) and V is the volume of culture media (L).

The number of mole of CO₂ supply (n_{CO_2-sup} , mol) can be determined from the following equation:

$$n_{CO_2-sup} = \frac{Qtp}{M},$$

where p is the CO₂ concentration, and M is the molar volume of CO₂ (mol L⁻¹) and is calculated by the following equation:

$$M = \frac{22.4 \times (273 + T)}{273}$$

where T is the culture temperature (°C).

The calculated CO₂ dissolution efficiency (η) in CRWP was obtained:

$$\eta = \frac{n_{CO_2-dis}}{n_{CO_2-sup}} = \frac{A(C^* - C)VM}{p} \quad (4)$$

According to Eq. (4), the CO₂ dissolution efficiency in CRWP without microalgae can be calculated. Fig. 3 shows the relationship of dissolved CO₂ concentration in media and CO₂ dissolution efficiency. The maximal mean relative error between the experimental

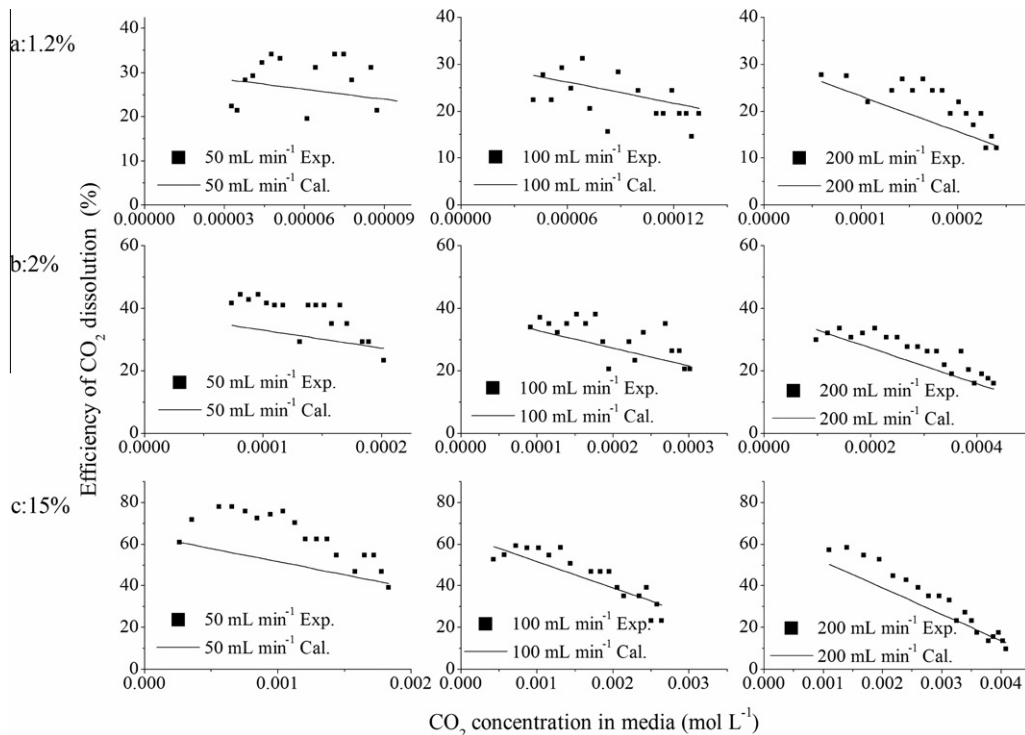
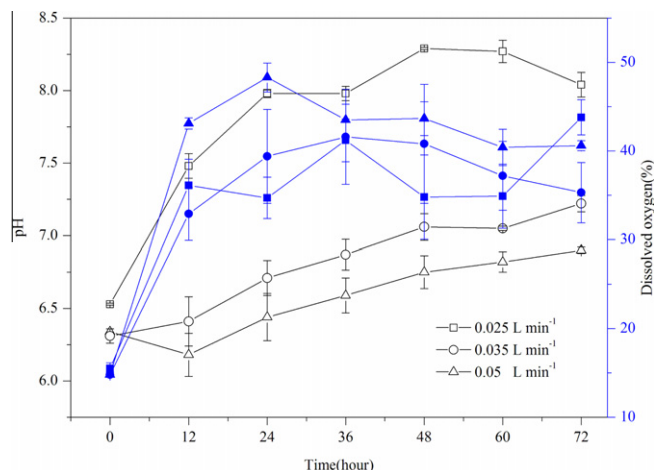


Fig. 3. Experimental and calculated values of CO₂ dissolution efficiency in CRWP. Exp. and Cal. represent experimental and calculated values, respectively. The experimental values were determined from the following equation: $\frac{C_{var}V}{n_{CO_2-sup}}$, where C_{var} was the variation of CO₂ concentration in media in five minutes (mol L⁻¹).

Table 1Microalgal productivity and CO₂ fixation efficiency with different gas flow rates.

Gas flow rate (L min ⁻¹)	Microalgal productivity (g d ⁻¹ L ⁻¹)	CO ₂ fixation efficiency (%) ^a
0.025	0.2349 ± 0.0168	53
0.035	0.3496 ± 0.0117	56
0.05	0.3219 ± 0.0109	36

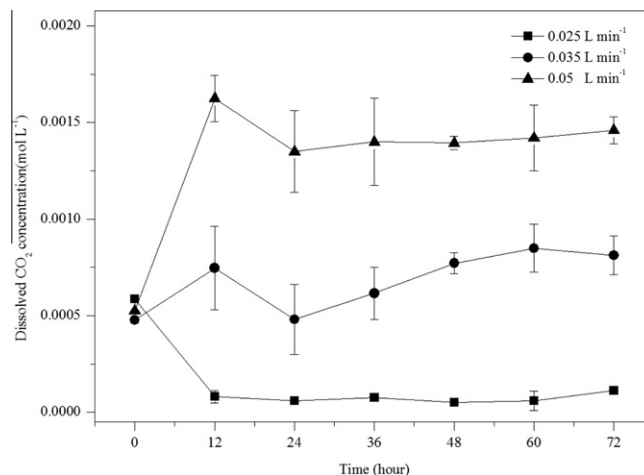
^a The CO₂ fixation efficiency was calculated as follows: $\frac{0.5PV}{12 \times 60 \times 24 \times \rho_{CO_2} \times \eta_{CO_2}} \times 100\%$, where 0.5, 12, 60 and 24 represent carbon percentage content in dried microalgae, carbon molecular weight, minutes in an hour and hours in a day, respectively.

**Fig. 4.** The pH values and DO levels of culture in CRWP. Open and closed symbols represent pH values and DO levels of culture, respectively.

and calculated values of dissolution efficiency was 14%, which showed a goodness of fit of the equation. As shown in Eq. (4), the efficiency of CO₂ dissolution in the CRWP aerated with certain CO₂ concentration had no relation with gas sparging rate and may be determined by the characteristics of CRWP, including the structure of photobioreactor, liquid velocity, mixing time and temperature, which influenced the overall volumetric mass transfer coefficient and hydrodynamics characteristics (Ugwu et al., 2008). The maximal CO₂ dissolution efficiency of media in CRWP aerated with 15% CO₂ was 64% according to Eq. (4), which was the maximal CO₂ fixation efficiency by microalgae under continuous CO₂ supply because microalgae can only utilize dissolved CO₂ and change it into biomass.

3.2. Microalgal productivity and CO₂ fixation efficiency with continuous gas sparging

The microalgal productivity and CO₂ fixation efficiency in CRWP under the concentration of 15% CO₂ with different gas flow rates are shown in Table 1. The changes of pH values and DO levels of culture during microalgal growth are shown in Fig. 4. As shown in Fig. 4, for 0.035 and 0.05 L min⁻¹, the pH values of culture ranged from 6.18 ± 0.14 to 7.22 ± 0.05, which did not inhibit microalgal growth (Hodaifa et al., 2010), and the levels of DO varied from 14.8 ± 0.1% to 48.3 ± 1.6% during cultivation. Although the oxygen build-up in photobioreactors would lead to photorespiration and even photo-oxidation with high levels of irradiance (Douskova et al., 2009; Miron et al., 1999), the competitive inhibition of ribulose-1,5-bisphosphate carboxylase by oxygen gradually became weak with the increase of concentration of CO₂ when oxygen level ranged from 20% to 50% (Aiba, 1982). In the present study, because the microalgal culture was aerated with high concentration of CO₂ (15% CO₂), and illuminated with low light intensity (100 μmol m⁻²

**Fig. 5.** Dissolved CO₂ concentrations in culture with different gas sparging rates.

s⁻¹), the inhibition action of oxygen accumulation to microalgal growth in the CRWP did not be found. Therefore, the culture pH and DO level in CRWP did not inhibit microalgal growth with the gas sparging rates of 0.035 and 0.05 L min⁻¹. Fig. 5 gives the change of dissolved CO₂ concentration in culture aerated with different gas sparging rates. As shown in Fig. 5, the average dissolved CO₂ concentrations in microalgal culture aerated with the gas sparging rates of 0.025, 0.035 and 0.05 L min⁻¹ from 12 to 72 h were 73, 713 and 1441 μmol L⁻¹, respectively. The microalgal productivity in CRWP aerated with the gas sparging rate of 0.025 L min⁻¹ was obviously lower than that of other gas flow rates, which showed that the dissolved CO₂ amounts cannot satisfy microalgal growth. Weissman et al. (1988) observed that CO₂ concentration in bulk liquid of at least 65 μmol L⁻¹ and pH 8.5 were required for optimal productivity of some microalgae, but in the present study, CO₂ concentration of 73 μmol L⁻¹ in media and pH 8 (shown in Fig. 4) cannot meet microalgal normal growth. The difference may be determined by algal species. For 0.035 and 0.05 L min⁻¹, with the increase of gas flow rate, the concentration of dissolved CO₂ in culture increased and the CO₂ fixation efficiency decreased, which indicated that much of CO₂ supply was wasted with continuous supply. According to Eq. (4), the maximal microalgal productivity in CRWP aerated with gas flow rates of 0.035 and 0.05 L min⁻¹ should be 0.3970 and 0.5670 g d⁻¹ L⁻¹, respectively, but the experimental microalgal productivity was 0.3496 and 0.3219 g d⁻¹ L⁻¹, respectively, which also indicated that some of supplied CO₂ was wasted.

3.3. CO₂ fixation efficiency by microalgae with intermittent gas sparging

With continuous gas sparging rates of 0.035 and 0.05 L min⁻¹, the fixation efficiency was 56% and 36%, respectively. According to Eq. (4), even the dissolved CO₂ was completely fixed by microalgae, the maximal CO₂ fixation efficiency in CRWP aerated with 15% CO₂ was 64%, which indicated there were much CO₂ escaped from CRWP. The method to further improve CO₂ fixation efficiency in CRWP was to increase the length of raceway, which would prolong the residence time of CO₂ in CRWP, or supply CO₂ intermittently. In this work, the later was further studied. In order to reach the maximal CO₂ fixation efficiency of 100%, the amounts of supplied CO₂ should completely dissolve in culture, and meanwhile, the dissolved CO₂ met microalgal growth need. Therefore, the relation between the gas sparging time (t_{aer}), the stopping time (t_{sto}), CO₂ dissolution rate in gas sparging (v_{dis}) and stopping (v_{res})

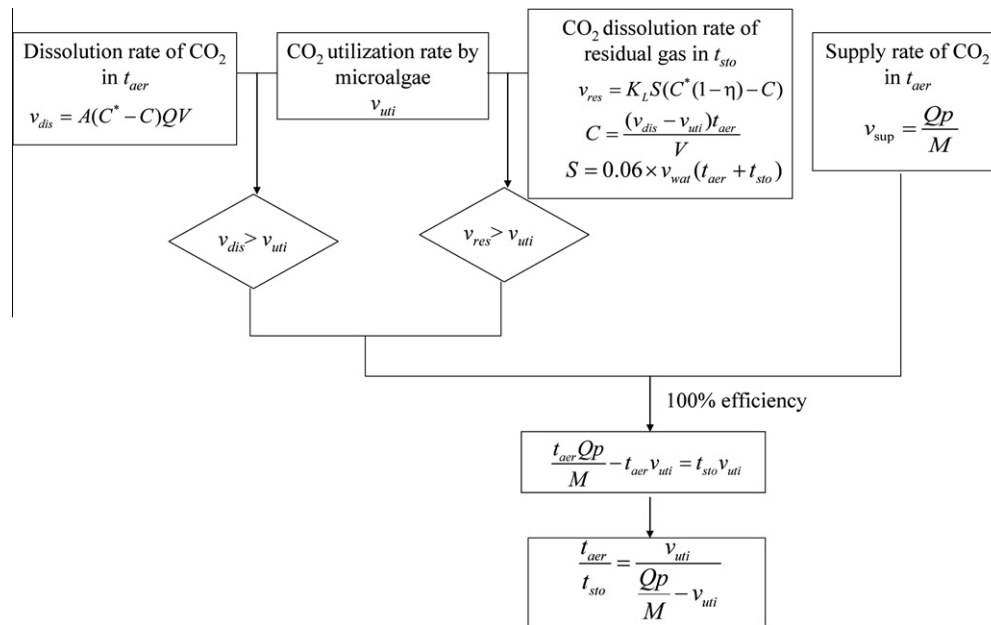


Fig. 6. The calculation model of intermittent time (0.06 is the width of each channel in the closed raceway pond).

time and the rate of CO₂ utilization (v_{uti}) by microalgae should be studied. In the stopping time, the remaining CO₂ (including CO₂ in culture and residual gas) should be enough for microalgal growth, and CO₂ in residual gas could dissolve completely in culture in t_{sto} . The dissolution rate of CO₂ in residual gas (v_{res}) can be expressed as follows:

$$v_{res} = K_L S(C^*(1 - \eta) - C),$$

where K_L is equal to 0.0096 m min⁻¹ (Shah et al., 1982) and S is the surface area of residual gas bubble in CRWP.

In the gas sparging time, CO₂ dissolution rate (v_{dis}) can be calculated from Eq. (3), and the average microalgal productivity (P) can be used to calculate the rate of CO₂ utilization (v_{uti}) by microalgae. Fig. 6 gives the calculation model of intermittent time. For example, under the gas flow rate of 0.035 L min⁻¹ with 15% CO₂ at 24 °C, the average microalgal productivity was 0.3496 g d⁻¹ L⁻¹ with continuous gas sparging (shown in Table 1). Through calculating, the dissolution rate of CO₂ (v_{dis}) was greater than the utilization rate of CO₂ by microalgae (v_{uti}) in the gas sparging time, and in the stopping time, v_{res} was far greater than v_{uti} , which indicated that the utilization rate of CO₂ by microalgae was the rate-limiting step for CO₂ absorption and the stopping time (t_{sto}) was determined by the utilization rate of CO₂ by microalgae (v_{uti}) in the intermittent operation. Therefore, the amounts of CO₂ supply in the gas sparging time should be equal to that of CO₂ of microalgal utilization in the gas sparging and stopping time, which gives:

$$\frac{t_{aer} Qp}{M} = t_{aer} v_{uti} + t_{sto} v_{uti};$$

For the gas sparging rate of 0.035 L min⁻¹ in CRWP, the proportion of t_{aer} to t_{sto} was closed to 1. The batch microalgal culture with the gas sparging time of 10 s and the stopping time of 10 s was investigated. After five days of cultivation, the average microalgal productivity was 0.2935 ± 0.0023 g d⁻¹ L⁻¹, which indicated that the CO₂ fixation efficiency by microalgae reached 95% in CRWP with intermittent gas sparging. Compared with continuous supply of CO₂, the intermittent gas sparging decreased the amounts of CO₂ supply and meanwhile the amounts met microalgal growth need, which improved the fixation efficiency. The values of t_{aer} and t_{sto} should be further analyzed. Under intermittent gas sparging, with the utilization of CO₂ by microalgae, the supplied CO₂ should com-

pletely dissolve in culture before running out of the CRWP, which meant that t_{aer} should be shorter than the residence time of CO₂ in CRWP with continuous supply, and on the other hand, the shorter of t_{aer} and t_{sto} , the faster of dissolution rate of CO₂ in culture, which benefitted the CO₂ fixation by microalgae. However, the microalgal productivity should be determined previously. Microalgal productivity can be predicted depending on light transfer with rich nutrition in medium (Fouchard et al., 2009; Yun and Park, 2003). Through the relationship between microalgal productivity, light distribution and CO₂ dissolution rate, the proportion of t_{aer} to t_{sto} can be calculated.

4. Conclusions

In this work, the CO₂ mass transfer, dissolution efficiency and CO₂ fixation efficiency by microalgae in the closed raceway pond were investigated. The dynamic equation for CO₂ dissolution and the method of improving CO₂ fixation efficiency by microalgae were developed. The experimental results showed that the developed model provided the guide for the improvement of CO₂ fixation efficiency by microalgae. Through the developed model, under intermittent operation (10 s:10 s), the fixation efficiency reached 95%. The concrete time of intermittent operation with different CO₂ partial pressures and gas flow rates should be further studied.

Acknowledgements

This study was funded by National Natural Science Foundation (Nos. 31101918 and 41276143) and Shandong Province Science & Technology Development Project (Nos. 2011GHY11531 and 2010GHY10504) and Natural Science Foundation of Shandong Province (Nos. Y2008F45 and ZR2012CL23).

References

- Aiba, S., 1982. Growth kinetics of photosynthetic microorganisms. Adv. Biochem. Eng. 23, 85–156.
- Becker, E.W., 1994. Microalgae Biotechnology and Microbiology. Cambridge University Press.
- Campbell, P.K., Beer, T., Batten, D., 2011. Life cycle assessment of biodiesel production from microalgae in ponds. Bioresour. Technol. 102, 50–56.

- Carvalho, A.P., Meireles, L.A., Malcata, F.X., 2006. Microalgal reactors: a review of enclosed system designs and performances. *Biotechnol. Prog.* 22, 1490–1506.
- Cheng, L., Zhang, L., Chen, H., Gao, C., 2006. Carbon dioxide removal from air by microalgae cultured in a membrane-photobioreactor. *Sep. Purif. Technol.* 50, 324–329.
- Chiu, S.-Y., Kao, C.-Y., Chen, C.-H., Kuan, T.-C., Ong, S.-C., Lin, C.-S., 2008. Reduction of CO₂ by a high-density culture of *Chlorella* sp. in a semicontinuous photobioreactor. *Bioresour. Technol.* 99, 3389–3396.
- de Morais, M.G., Costa, J.A.V., 2007. Biofixation of carbon dioxide by *Spirulina* sp. and *Scenedesmus obliquus* cultivated in a three-stage serial tubular photobioreactor. *J. Biotechnol.* 129, 439–445.
- Douskova, I., Doucha, J., Livansky, K., Machat, J., Novak, P., Umysova, D., Zachleder, V., Vitova, M., 2009. Simultaneous flue gas bioremediation and reduction of microalgal biomass production costs. *Appl. Microbiol. Biotechnol.* 82, 179–185.
- Fouchard, S., Pruvost, J., Degrenne, B., Titica, M., Legrand, J., 2009. Kinetic modeling of light limitation and sulfur deprivation effects in the induction of hydrogen production with *Chlamydomonas reinhardtii*: Part I. Model development and parameter identification. *Biotechnol. Bioeng.* 102, 232–245.
- Hodaifa, G., Martínez, M.E., Sánchez, S., 2010. Influence of pH on the culture of *Scenedesmus obliquus* in olive-mill wastewater. *Biotechnol. Bioprocess Eng.* 14, 854–860.
- Keffer, J.E., Kleinheinz, G.T., 2002. Use of *Chlorella vulgaris* for CO₂ mitigation in a photobioreactor. *J. Ind. Microbiol. Biotechnol.* 29, 275–280.
- Ketheesan, B., Nirmalakhandan, N., 2012. Feasibility of microalgal cultivation in a pilot-scale airlift-driven raceway reactor. *Bioresour. Technol.* 108, 196–202.
- Laws, E.A., Berning, J.L., 1991. A study of the energetics and economics of microalgal mass culture with the marine chlorophyte *Tetraselmis suecica*: implications for use of power plant stack gases. *Biotechnol. Bioeng.* 37, 936–947.
- Li, S.W., Luo, S.J., Guo, R.B., 2012. Influence of carbon dioxide concentration on microalgal growth in a bubble column photobioreactor. *Adv. Mater. Res.* 599, 137–140.
- Mata, T.M., Martins, A.A., Caetano, N.S., 2010. Microalgae for biodiesel production and other applications: a review. *Renew. Sust. Energy Rev.* 14, 217–232.
- Milledge, J.J., 2010. Commercial application of microalgae other than as biofuels: a brief review. *Rev. Environ. Sci. Biotechnol.* 10, 31–41.
- Miron, A.S., Gomez, A.C., Camacho, F.G., Grima, E.M., Chisti, Y., 1999. Comparative evaluation of compact photobioreactors for large-scale monoculture of microalgae. *J. Biotechnol.* 70, 249–270.
- Putt, R., Singh, M., Chinnasamy, S., Das, K.C., 2011. An efficient system for carbonation of high-rate algae pond water to enhance CO₂ mass transfer. *Bioresour. Technol.* 102, 3240–3245.
- Shah, Y., Kelkar, B.G., Godbole, S., Deckwer, W.D., 1982. Design parameters estimations for bubble column reactors. *AIChE J.* 28, 353–379.
- Spolaore, P., Joannis-Cassan, C., Duran, E., Isambert, A., 2006. Commercial applications of microalgae. *J. Biosci. Bioeng.* 101, 87–96.
- Ugwu, C.U., Aoyagi, H., Uchiyama, H., 2008. Photobioreactors for mass cultivation of algae. *Bioresour. Technol.* 99, 4021–4028.
- Weissman, J.C., Goebel, R.P., Benemann, J.R., 1988. Photobioreactor design: mixing, carbon utilization, and oxygen accumulation. *Biotechnol. Bioeng.* 31, 336–344.
- Weissman, J.C., Goebel, R.P., 1985. Production of Liquid Fuels and Chemicals by Microalgae. Solar Energy Research Institute, Golden, Co., SERI/STR-231-2649.
- Yun, Y.S., Park, J.M., 2003. Kinetic modeling of the light-dependent photosynthetic activity of the green microalga *Chlorella vulgaris*. *Biotechnol. Bioeng.* 83, 303–311.